

EVIDENCE FOR FREE RADICAL SCAVENGING ACTIVITY OF ASHWAGANDHA ROOT POWDER IN MICE

SUNANDA PANDA AND ANAND KAR*

*School of Life Sciences, D. A. University,
Vigyan Bhawan, Khandwa Road,
Indore - 452 001*

(Received on September 20, 1996)

Abstract: The effects of Ashwagandha root powder (0.7 and 1.4 g/kg body weight/day), administered for 15 and 30 days, was investigated on lipid peroxidation (LPO), superoxide dismutase (SOD) and catalase (CAT) activities in mice. While 15 days treatment with Ashwagandha root powder did not produce any significant change, 30 days treatment produced a significant decrease in LPO, and an increase in both SOD and CAT. Our findings indicate that Ashwagandha root powder possesses free radical scavenging activity, which may be responsible for its pharmacological effects.

Key words: Ashwagandha
catalase

superoxide dismutase
lipid peroxidation

INTRODUCTION

Free radical induced lipid peroxidation has gained much importance, because of its involvement in several pathological conditions such as atherosclerosis, ischemia, liver disorder, neural disorder, metal toxicity and pesticide toxicity (1-3). Lipid peroxidation (LPO) involves the formation of lipid radicals, oxidation of unsaturated lipids and the eventual destruction of membrane lipids producing a variety of breakdown products and deleterious effects. These deleterious processes can be prevented by reducing or destroying the formation of free radicals which are continuously formed in a biological system. In normal tissues, endogenous enzymes are there that counteract the free radicals, superoxide dismutase (SOD) and catalase (CAT) being the most common ones.

However, clinically other antioxidants such as vitamin E and vitamin C are also used. Among the herbal preparations only limited trials have been done and not much is known (2,4). We studied the antioxidant activity of the root powder of a subtropical undershrub, Ashwagandha (*Withania somnifera*), that is well known for its medicinal value (5).

METHOD

Fifteen adults healthy colony bred 'Swiss albino' male mice, weighing 30-35 g (approximate age 3 months) were maintained in controlled room temperature ($27 \pm 1^\circ\text{C}$) and light cycle (14 h light and 10 h dark), with food (obtained from Hindustan Lever Ltd., India) and water *ad libitum*. After an acclimatization of 7 days, mice were divided into three groups of five

* Corresponding Author

each. The initial body weight of each was recorded.

Ashwagandha treatment: Ashwagandha root, dried powder (with 1.75% Withanolides) supplied by Kisalaya Pharmaceuticals Pvt. Ltd., Indore, India was used to prepare suspensions of 20 mg/0.1 ml and 40 mg/0.1 ml concentrations using water : tween 80, (9 : 1), for the treatment.

Group I received equivalent amount of the suspending agent and served as control. Group II and Group III received 0.7 g and 1.4 g of Ashwagandha suspension per kg body weight/day respectively. Treatment was given orally by gastric intubation, every day between 10–11 h and continued for 30 days. On the last day, final body weight of each animal was recorded. Animals were then sacrificed, liver of each animal was removed, cleaned twice in phosphate buffered saline (pH 7.4) and immediately processed for biochemical estimations.

LPO was studied by the method of Ohkawa *et al* (6) followed earlier in our laboratory. In brief, liver was homogenised in 10% W/V ice cold phosphate buffer (0.1 M, pH 7.4) using Potter-Elvehjem teflon homogenizer. The homogenate was centrifuged at 2,000 g at 4°C for 30 min and supernatant was used for subsequent assay procedure. LPO was determined by thiobarbituric acid (TBA) reaction with malondialdehyde (MDA), a product formed due to peroxidation of lipids. The LPO was expressed as nM of MDA formed per h per mg protein. SOD activity was determined by the method of Marklund and Marklund (7) and CAT activity by the method of Aebi

(8). The values were expressed as μmol of hydrogen peroxide decomposed/min/mg protein.

These experiments were repeated with another 15 albino mice. However, this time treatment was for 15 days only in order to evaluate the effectiveness of the short term drug therapy.

Data have been expressed as the mean \pm standard error of the mean (SEM). For statistical evaluation, Students 't' test was used.

RESULTS AND DISCUSSION

Ashwagandha suspension decreased LPO significantly ($P < 0.001$ for both high and low doses). In contrast, it significantly increased the SOD ($P < 0.05$ for low dose and $P < 0.001$ for high dose) and CAT ($P < 0.05$ for low dose and $P < 0.001$ for high dose) compared to the respective control values (Table I). These results clearly demonstrate a drug induced inhibition in lipid peroxidative process, indicating protective effect of the Ashwagandha root powder. Administration of this drug inhibited the formation of hepatic malondialdehyde

TABLE I: Values (mean \pm SEM) of LPO (nM of MDA/h/mg protein), SOD (Units/mg protein) and CAT (μM of H_2O_2 decomposed/min/mg protein) activities in control and Ashwagandha treated (0.7 and 1.4 g/kg body weight/day for 30 days) mice.

Groups	LPO	SOD	CAT
Control	0.48 \pm 0.02	5.92 \pm 0.18	55.19 \pm 1.86
Treated (0.7 g)	0.35 \pm 0.02***	7.56 \pm 0.51*	67.15 \pm 2.21**
Treated (1.4 g)	0.25 \pm 0.01***	9.21 \pm 0.65***	71.32 \pm 1.5***

* $P < 0.05$; ** $P < 0.01$ and *** $P < 0.001$ compared to the respective control values.

significantly only when treatment was continued for 30 days suggesting that, the effectiveness of the drug depends on the duration of the treatment. Similar to our finding, protective effect with respect to LPO has also been reported for two other plants, *Rubia cordifolia* (2) and *Bacopa monniera* (4).

Hydrogen peroxide (H_2O_2) is continuously generated in large concentrations by the mitochondrial inner membrane's respiratory chain. Oxygen gets converted to superoxide anion radical (O_2^-) when it acquires an extra electron. Most of these radicals are destroyed by SOD. Superoxide anion (O_2^-) reacts with water to form H_2O_2 which in turn responsible for the highly reactive hydroxyl (OH^-) radicals that attack membrane fatty acids to initiate LPO and bring about necrotic changes in the tissue (9). Catalase appears to be the most

effective defensive agent against high concentrations of H_2O_2 . In the present study, interestingly, both SOD and CAT activities were increased by Ashwagandha, further supporting the defensive nature of the drug against free radical damage.

Thus Ashwagandha may act as an exogenous antioxidant. Since Ashwagandha is known for its aphrodisiac, psychotropic, antitumor and cardiogenic effects, some of these pharmacological effects may be due to free radical scavenging activity of the Ashwagandha root powder.

ACKNOWLEDGMENTS

Financial assistance from University Grant Commission, India (JRF to Ms. S. Panda) is acknowledged. Authors also thank P.K. Maiti, S.S. Chaurasia and P. Gupta who helped in various ways.

REFERENCES

1. Chaurasia SS, Gupta P, Maiti PK, Kar A. Free radical mediated membrane perturbation and inhibition of type-I iodothyronine 5'-monodeiodinase activity by lead and cadmium in rat liver homogenate. *Biochem Mol Biol Int* 1996; 39: 765-770.
2. Pandey S, Sharma M, Chaturvedy P, Tripathi YB. Protective effect of *Rubia cordifolia* on lipid peroxide formation in isolated rat liver homogenate. *Indian J Exp Biol* 1994; 3: 180-183.
3. Maiti PK, Kar A, Gupta P, Chaurasia SS. Loss of membrane integrity and inhibition of type-I iodothyronine 5'-monodeiodinase activity by fenvalerate in female mouse. *Biochem Biophys Res Commun* 1995; 214: 905-909.
4. Tripathi YB, Chaurasia S, Tripathi E, Upadhyay A, Dubey GP. *Bacopa monniera* Linn. as an antioxidant: Mechanism of action. *Indian J Exp Biol* 1996; 34: 523-526.
5. Tripathi AK, Shukla YN, Kumar S. Ashwagandha (*Withania somnifera*): A status report. *J Med Arom Plant Science* 1996; 18: 46-62.
6. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 1979; 95: 351-358.
7. Marklund S, Marklund G. Involvement of superoxide anion radical in autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur J Biochem* 1974; 47: 469-474.
8. Aebi H. Catalase. In: *Methods in Enzymatic Analysis*, Bergmeyer HU (ed), Vol. 3, New York, Academic Press, 1983: 276-286.
9. Gutteridge JMC, Richmond R, Halliwell B. Inhibition of the iron catalysed formation of hydroxyl radicals from superoxide and lipid peroxidation by desferrioxamine. *Biochem J* 1979; 184: 469-472.